

The resurgence of reference quality genomes

Michael Schatz

May 22, 2015

NYU Genomics Symposium



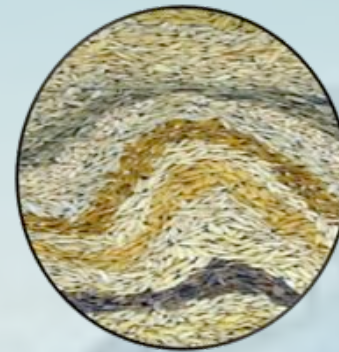
Schatzlab Overview



Human Genetics

Role of mutations in disease

Narzisi *et al.* (2014)
Iossifov *et al.* (2014)



Plant Biology

Genomes &
Transcriptomes

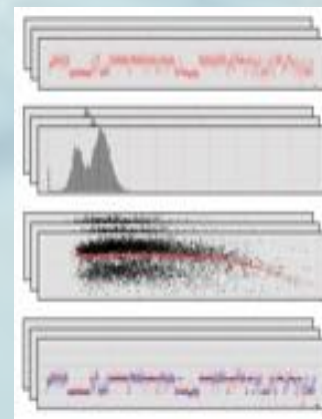
Schatz *et al.* (2014)
Ming *et al.* (2013)



Algorithmics & Systems Research

Ultra-large scale
biocomputing

Marcus *et al.* (2014)
Schatz *et al.* (2013)



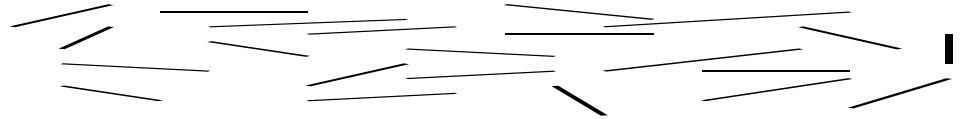
Single Cell & Single Molecule

CNVs, SVs, &
Cell Phylogenetics

Garvin *et al.* (2014)
Roberts *et al.* (2013)

Sequence Assembly Problem

1. Shear & Sequence DNA



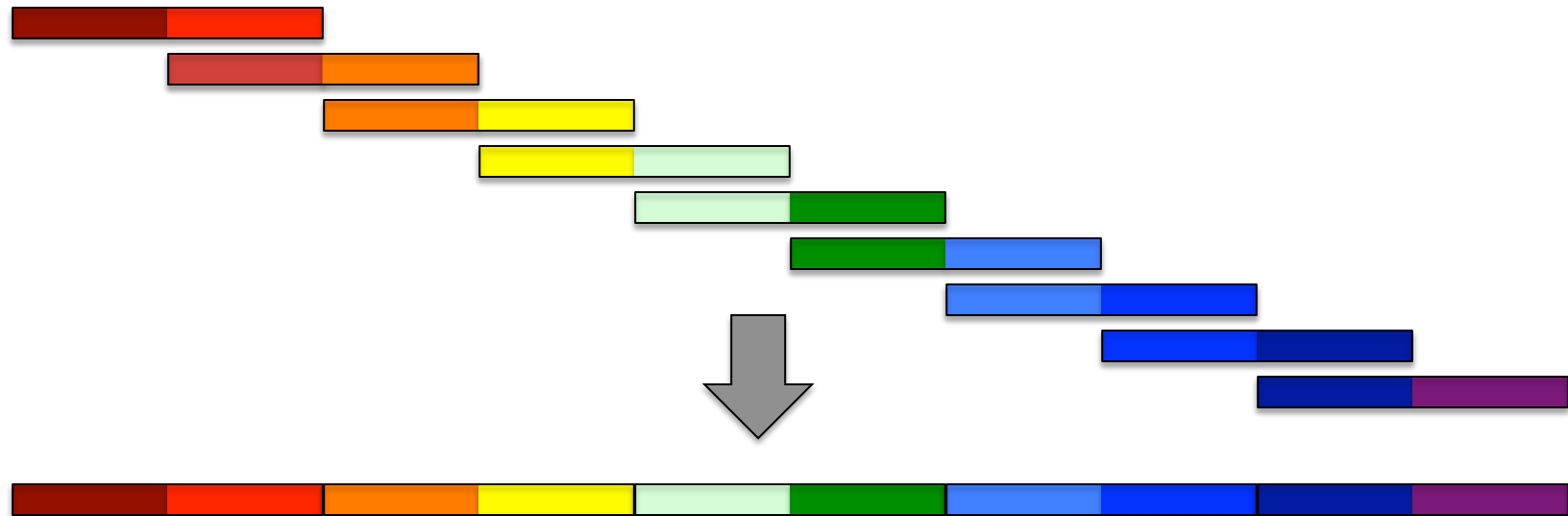
2. Construct assembly graph from overlapping reads

...AGCCTAGGGATGCGCGACACGT

GGATGCGCGACACGT CGCATATCCGGTTTGGT CAACCTCGGACGGAC

CAACCTCGGACGGACCTCAGCGAA...

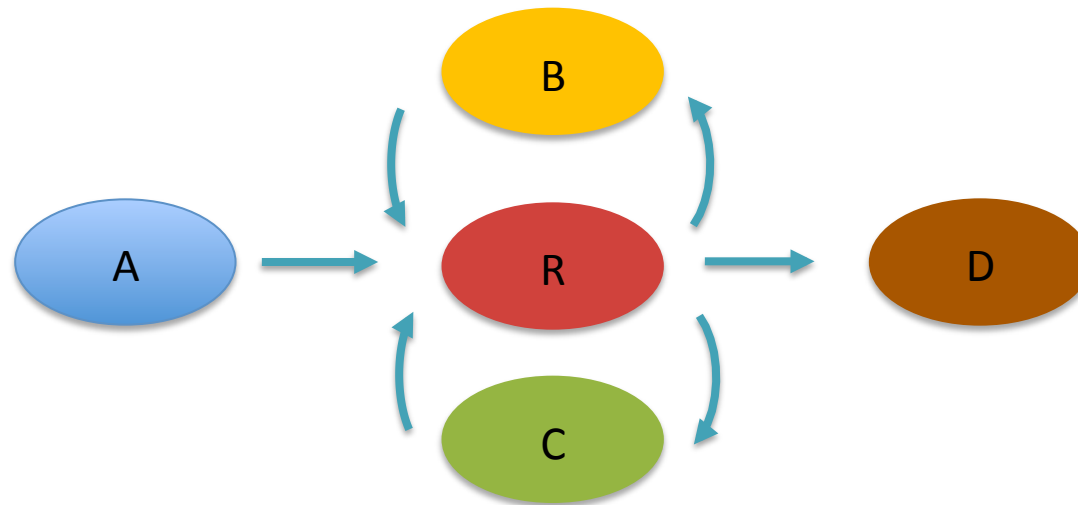
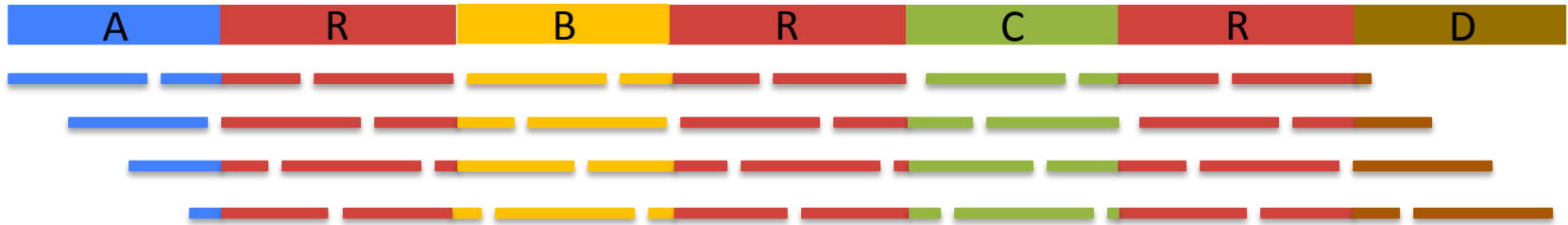
3. Simplify assembly graph



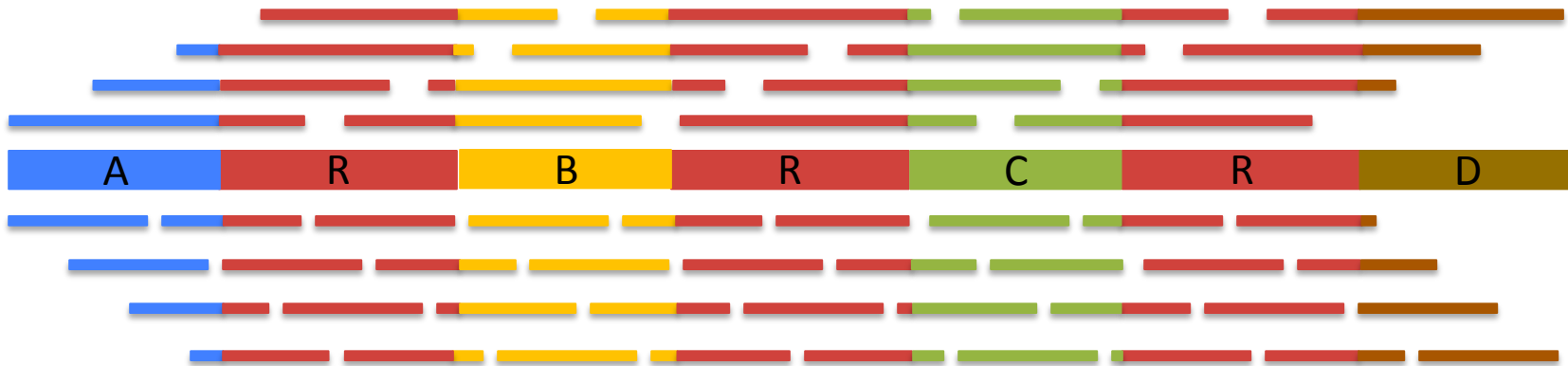
On Algorithmic Complexity of Biomolecular Sequence Assembly Problem

Narzisi, G, Mishra, B, Schatz, MC (2014) *Algorithms for Computational Biology*. Lecture Notes in Computer Science. Vol. 8542

Assembly Complexity



Assembly Complexity



The advantages of SMRT sequencing

Roberts, RJ, Carneiro, MO, Schatz, MC (2013) *Genome Biology*. 14:405

Genomics Arsenal in the Year 2015

Long Read Sequencing: De novo assembly, SV analysis, phasing

Illumina/Moleculo



(Kuleshov et al. 2014)

Pacific Biosciences



(Berlin et al, 2014)

Oxford Nanopore



(Quick et al, 2014)

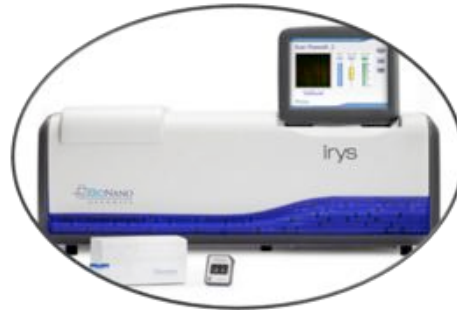
Long Span Sequencing: Chromosome Scaffolding, SV analysis, phasing

Molecular Barcoding



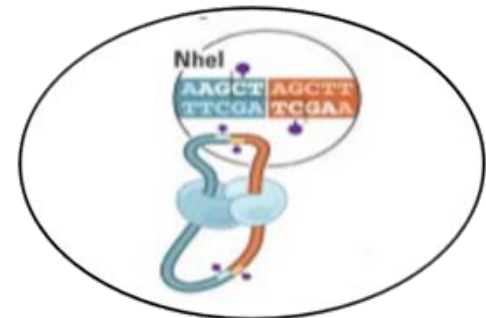
(10Xgenomics.com)

Optical Mapping



(Cao et al, 2014)

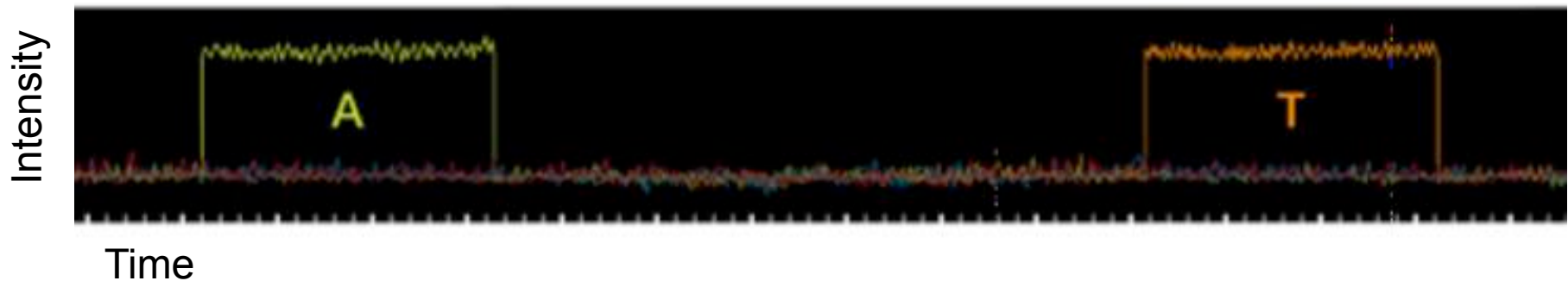
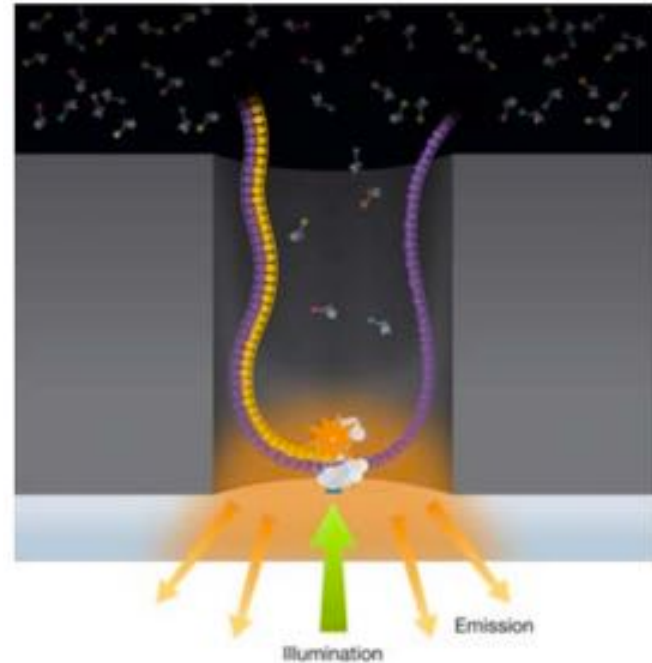
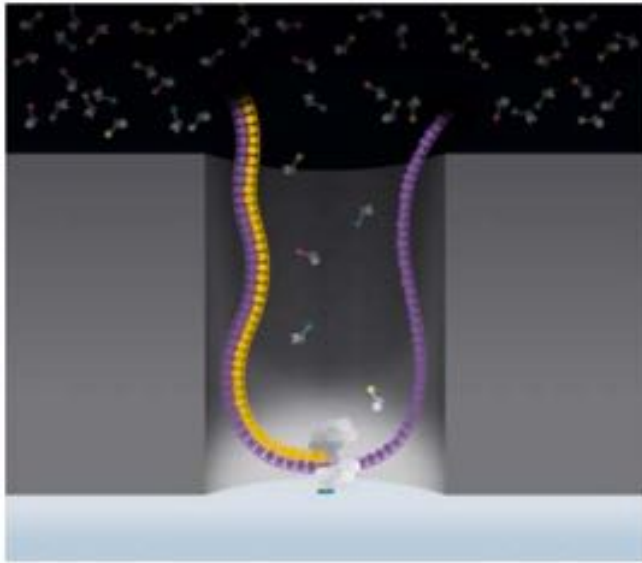
Chromatin Assays



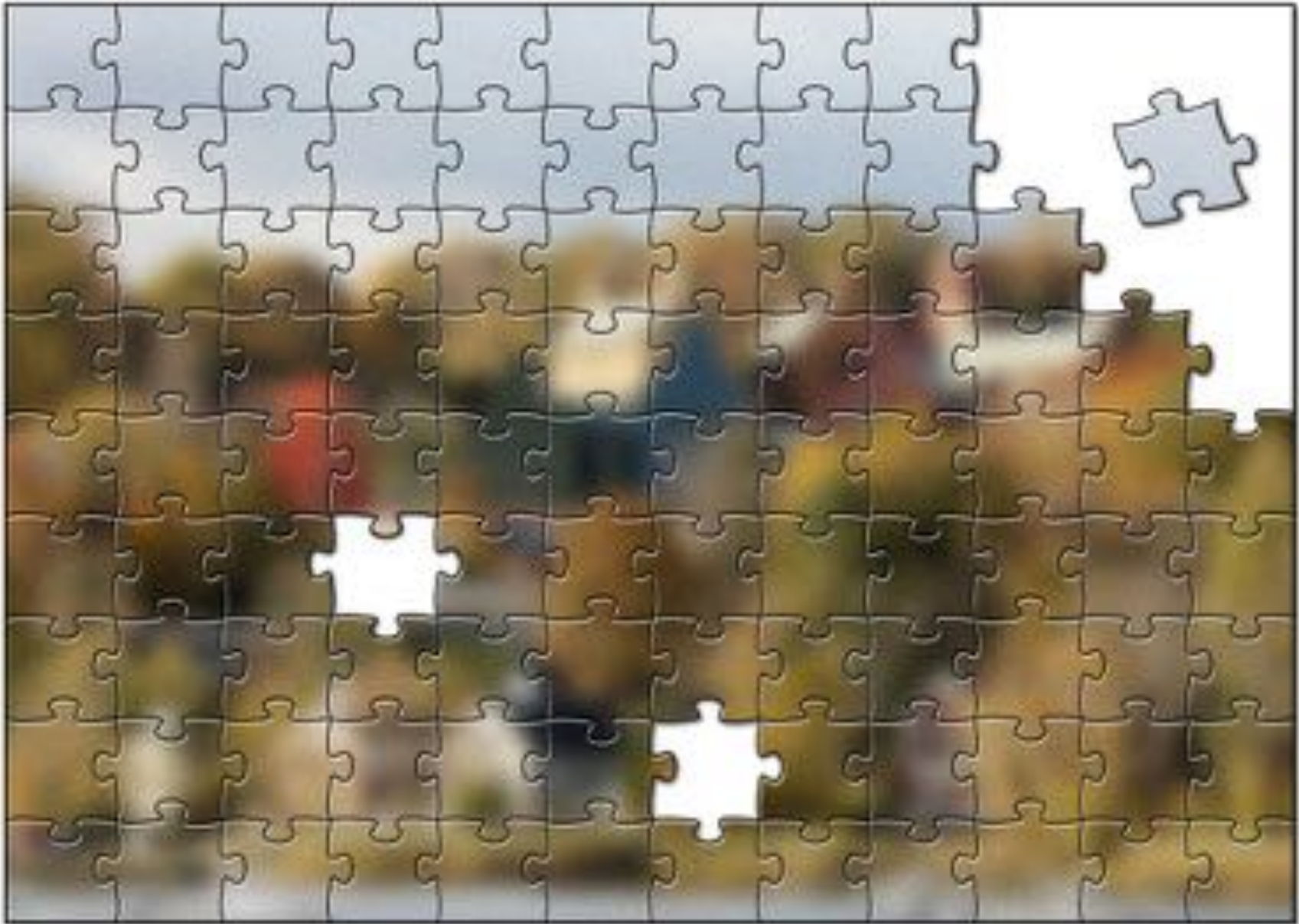
(Putnam et al, 2015)

PacBio SMRT Sequencing

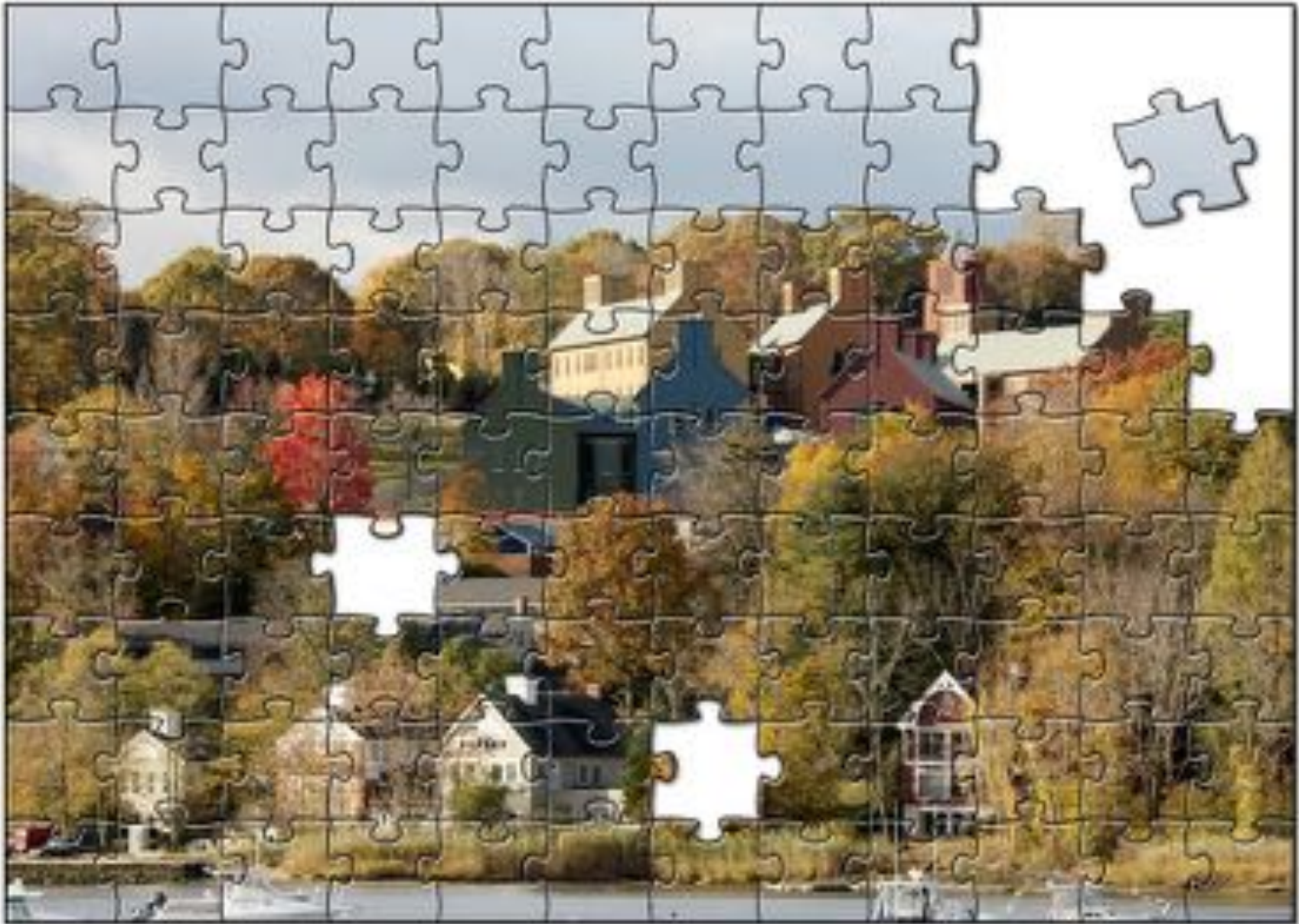
Imaging of fluorescently phospholinked labeled nucleotides as they are incorporated by a polymerase anchored to a Zero-Mode Waveguide (ZMW).



Single Molecule Sequences

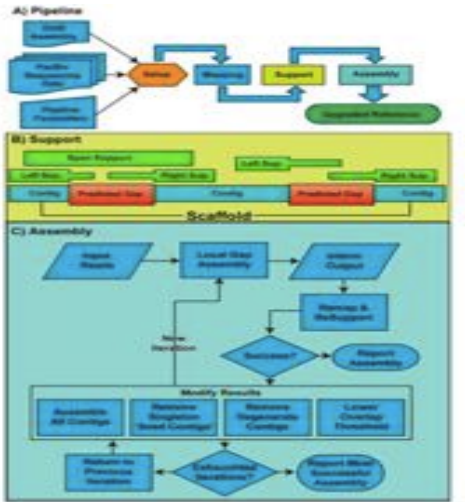


“Corrective Lens” for Sequencing



PacBio Assembly Algorithms

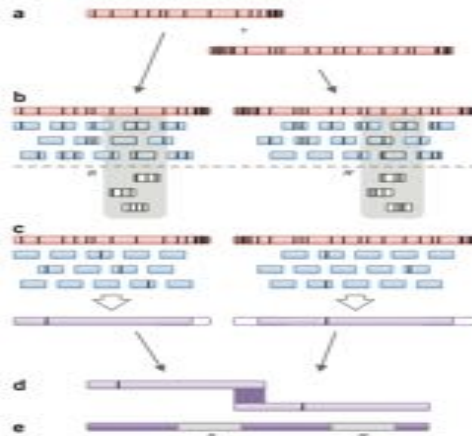
PBJelly



**Gap Filling
and Assembly Upgrade**

English *et al* (2012)
PLOS One. 7(11): e47768

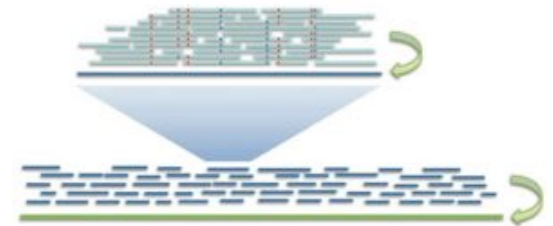
PacBioToCA & ECTools



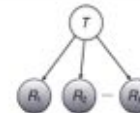
**Hybrid/PB-only Error
Correction**

Koren, Schatz, *et al* (2012)
Nature Biotechnology. 30:693–700

HGAP/MHAP & Quiver



$$\Pr(\mathbf{R} | T) = \prod_k \Pr(R_k | T)$$



Quiver Performance Results Comparison to Reference Genome (<i>M. ruber</i> ; 3.1 MB; SMRT® Cells)		
QV	Initial Assembly 43.4	Quiver Consensus 54.5
Accuracy	99.99540%	99.99964%
Differences	141	11

**PB-only Correction &
Polishing**

Chin *et al* (2013)
Nature Methods. 10:563–569

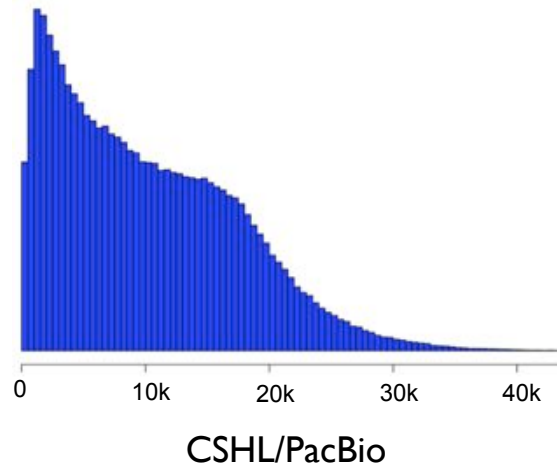
< 5x

PacBio Coverage

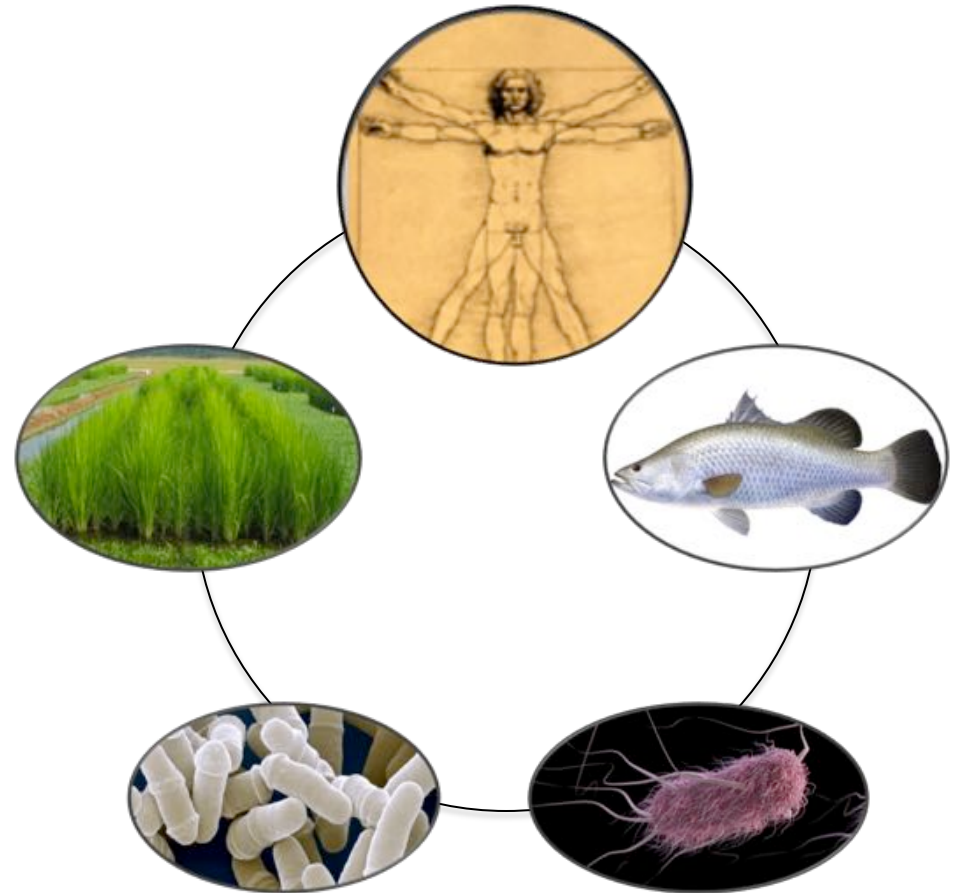
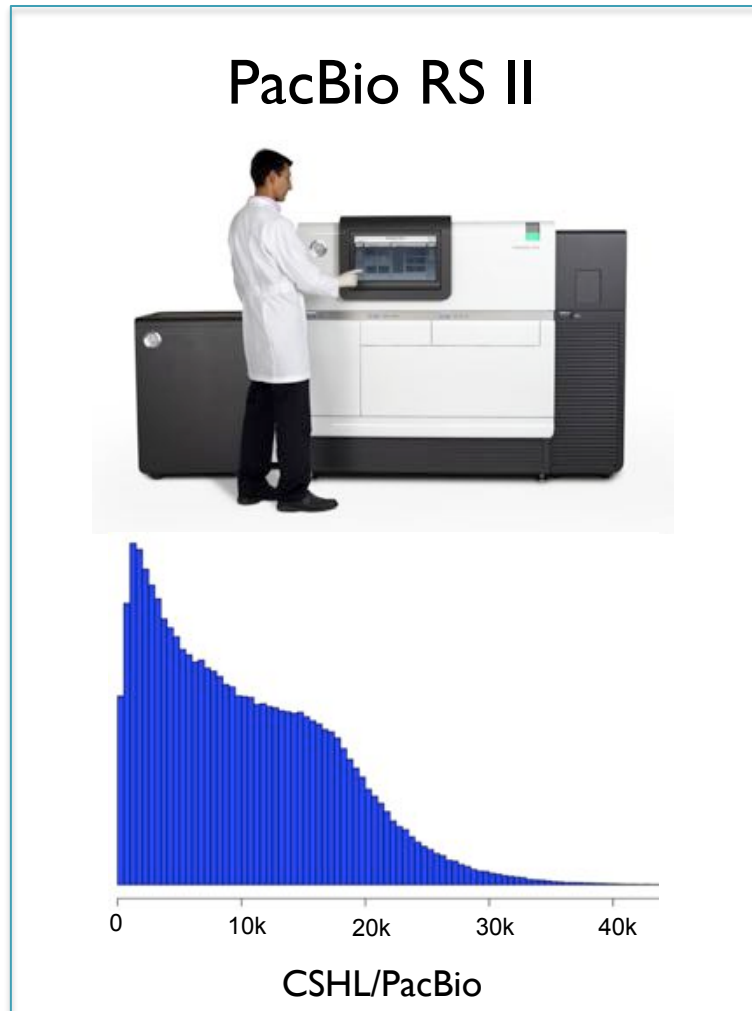
> 50x

3rd Gen Long Read Sequencing

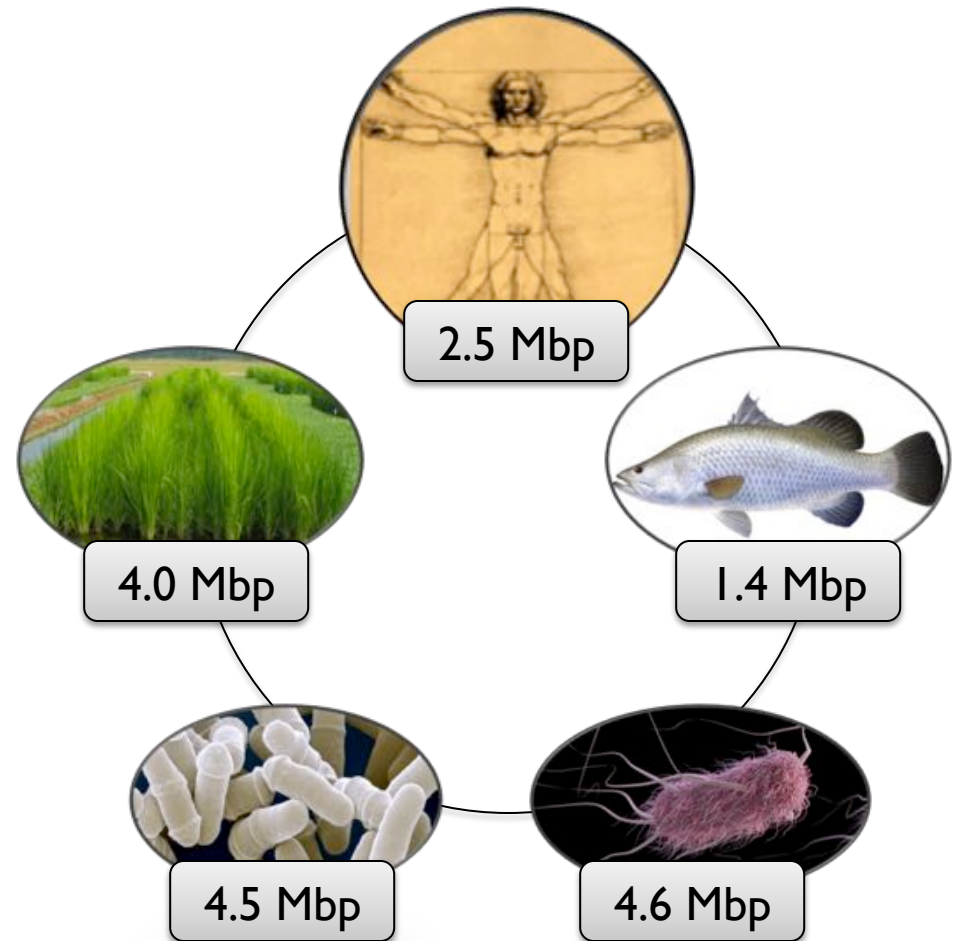
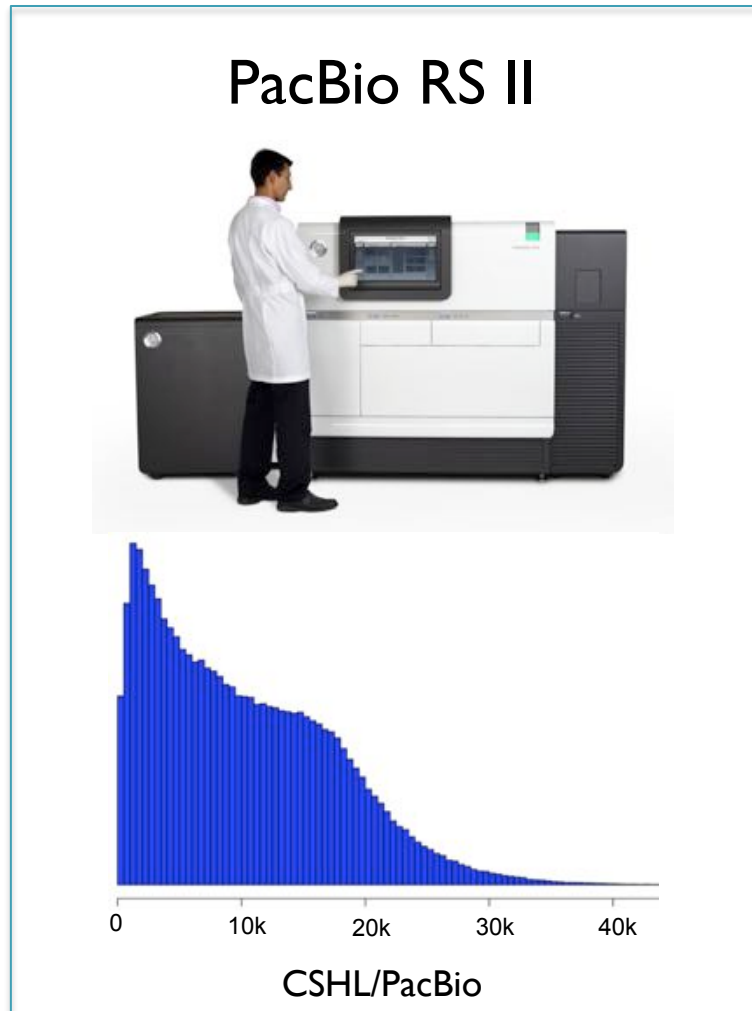
PacBio RS II



3rd Gen Long Read Sequencing

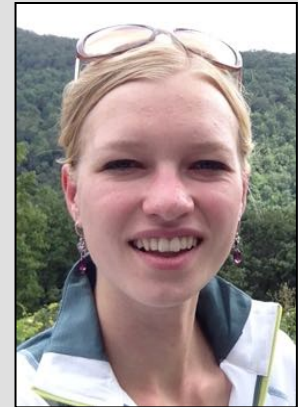


3rd Gen Long Read Sequencing

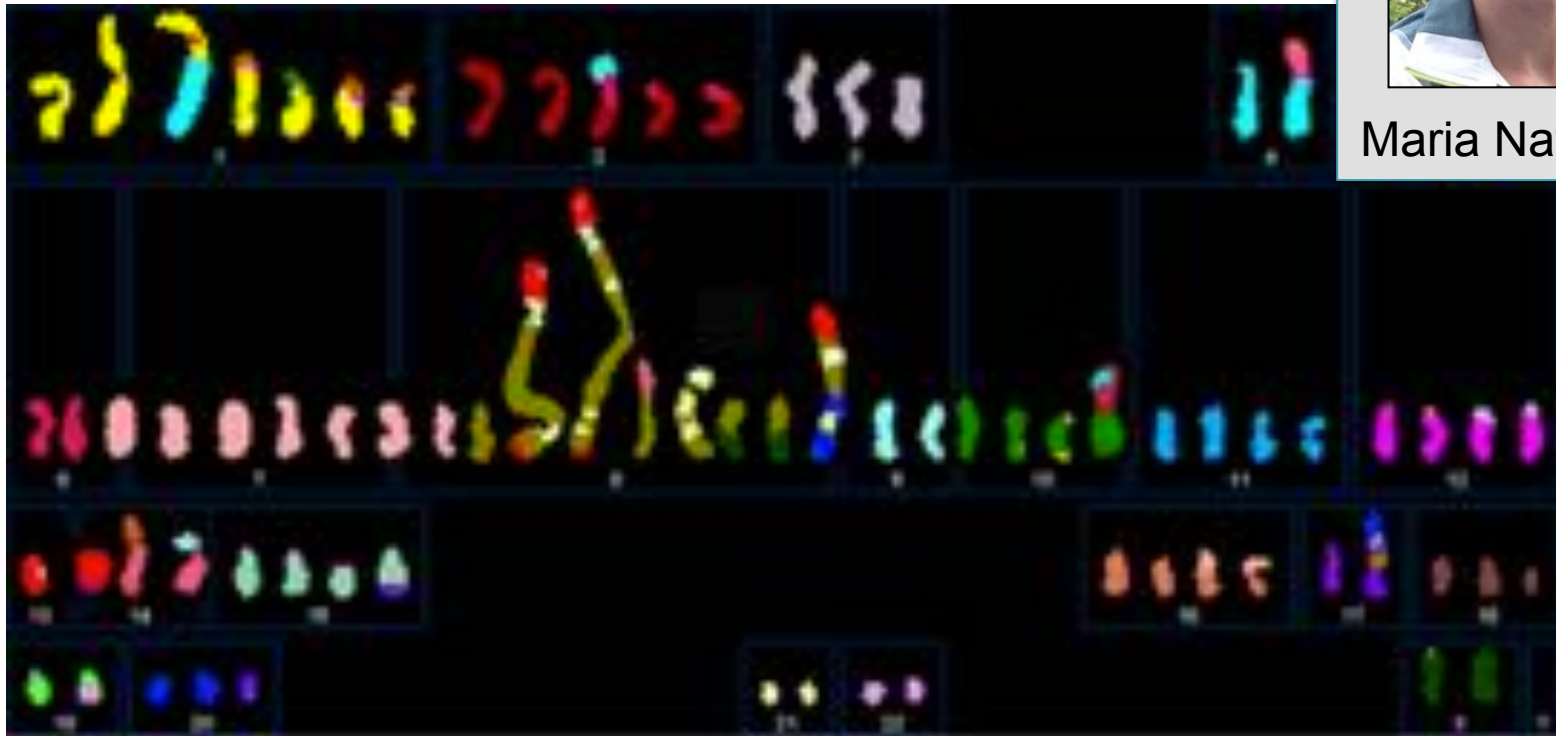


SK-BR-3

Most commonly used Her2-amplified breast cancer



Maria Nattestad

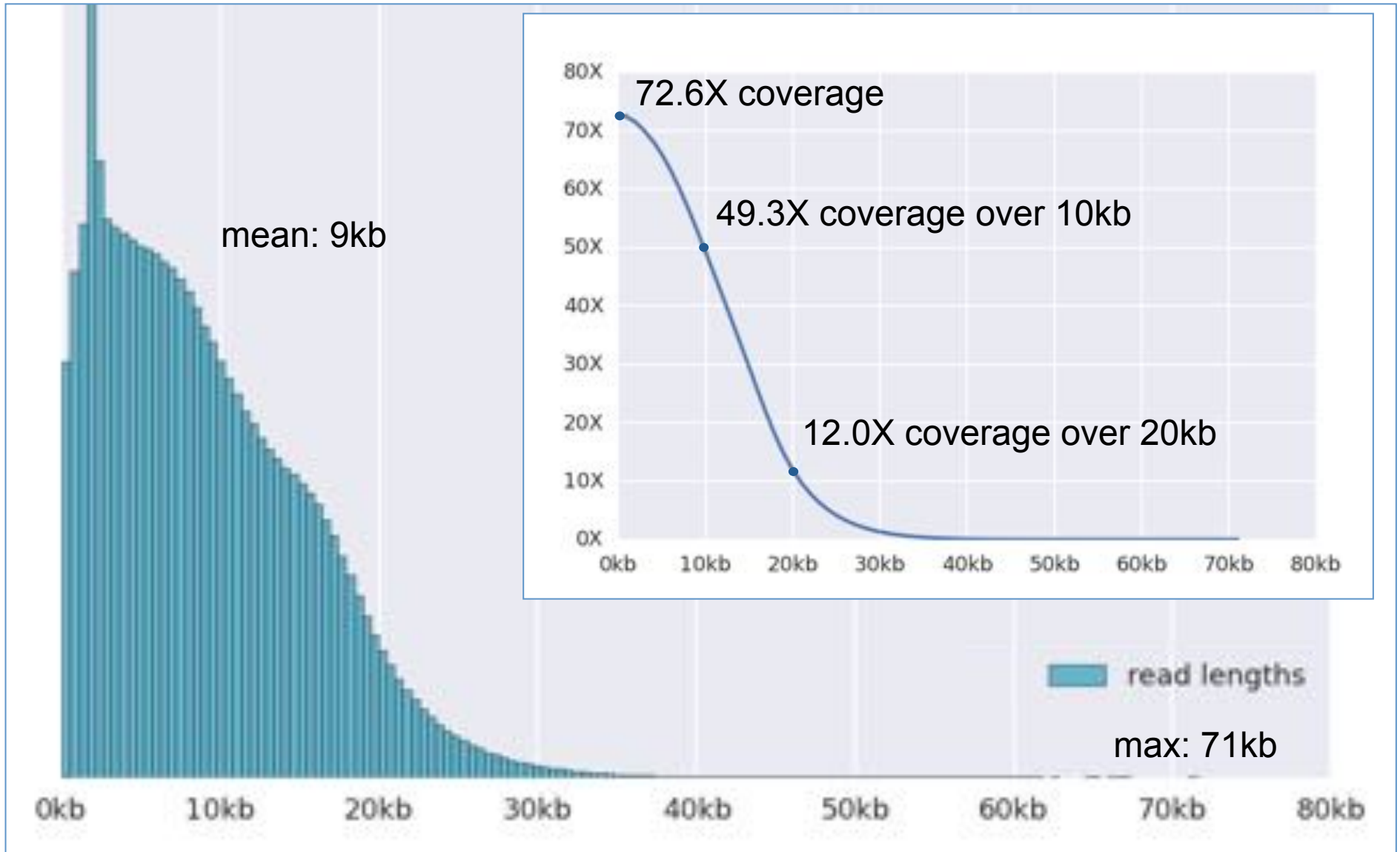


(Davidson et al, 2000)

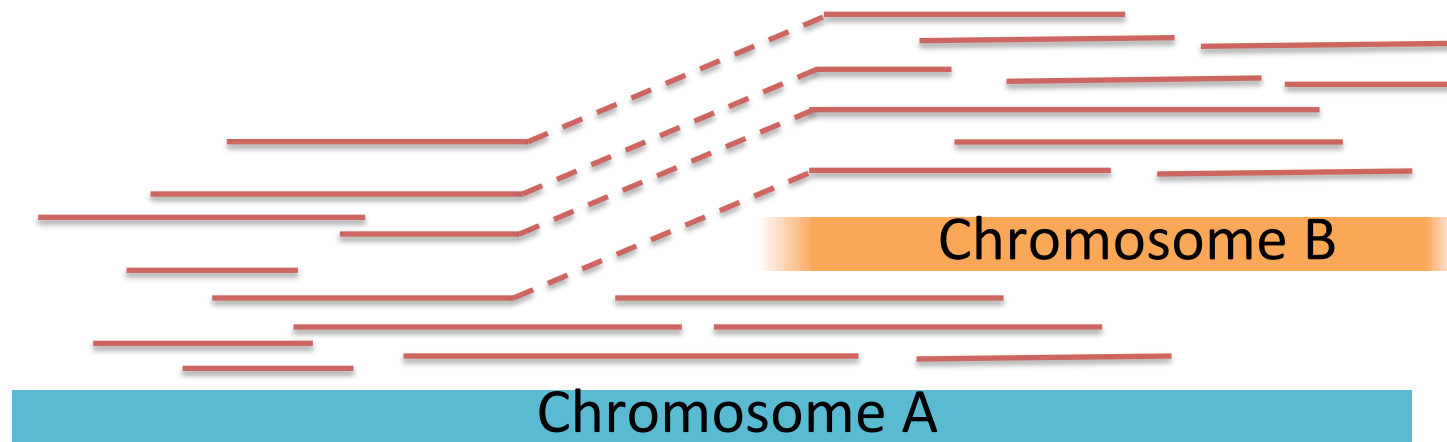
Can we resolve the complex structural variations, especially around Her2?

Ongoing collaboration between CSHL and OICR to *de novo* assemble the complete cell line genome with PacBio long reads

PacBio read length distribution



Structural variant discovery with long reads



1. Alignment-based split read analysis: Efficient capture of most events

BWA-MEM + Lumpy

2. Local assembly of regions of interest: In-depth analysis with *base-pair precision*

Localized HGAP + Celera Assembler + MUMmer

3. Whole genome assembly: In-depth analysis including *novel sequences*

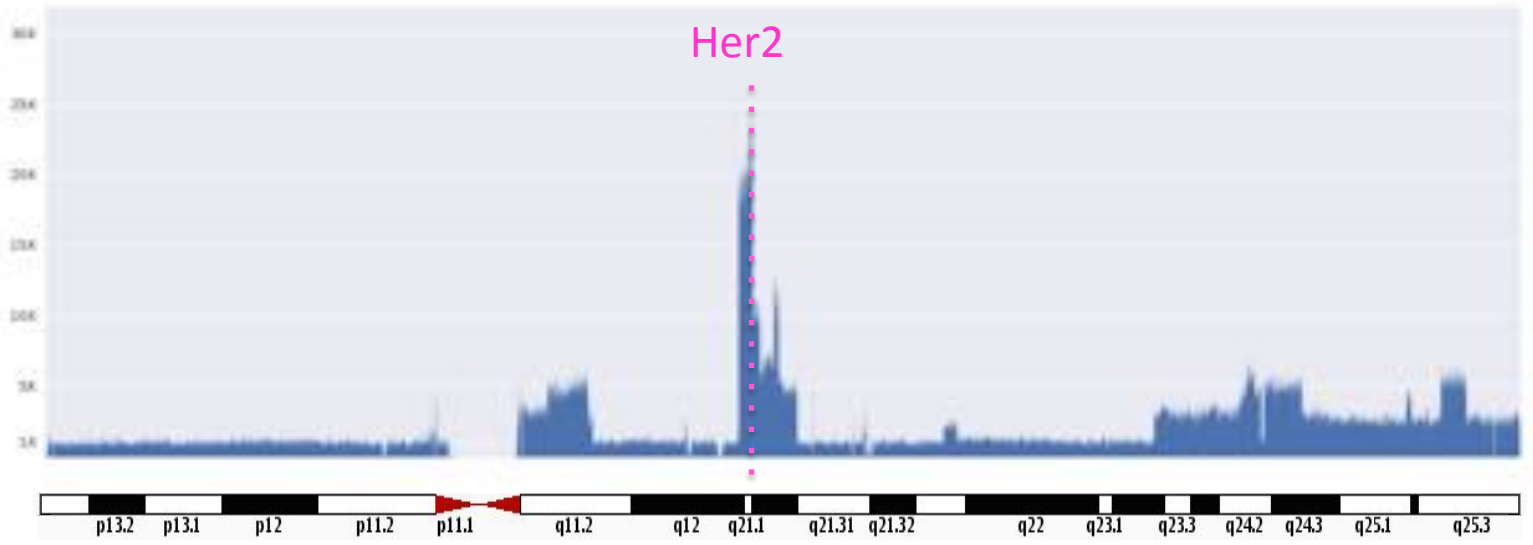
DNAnexus-enabled version of Falcon

Total Assembly: 2.64Gbp

Contig N50: 2.56 Mbp

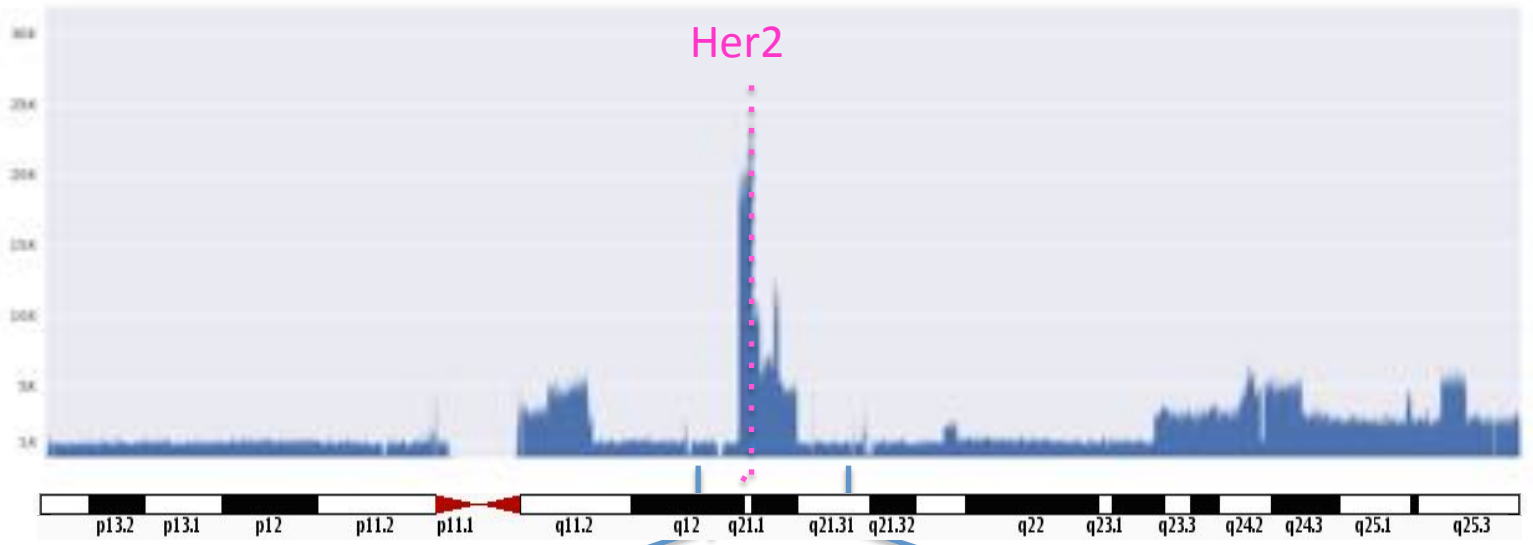
Max Contig: 23.5Mbp

PacBio

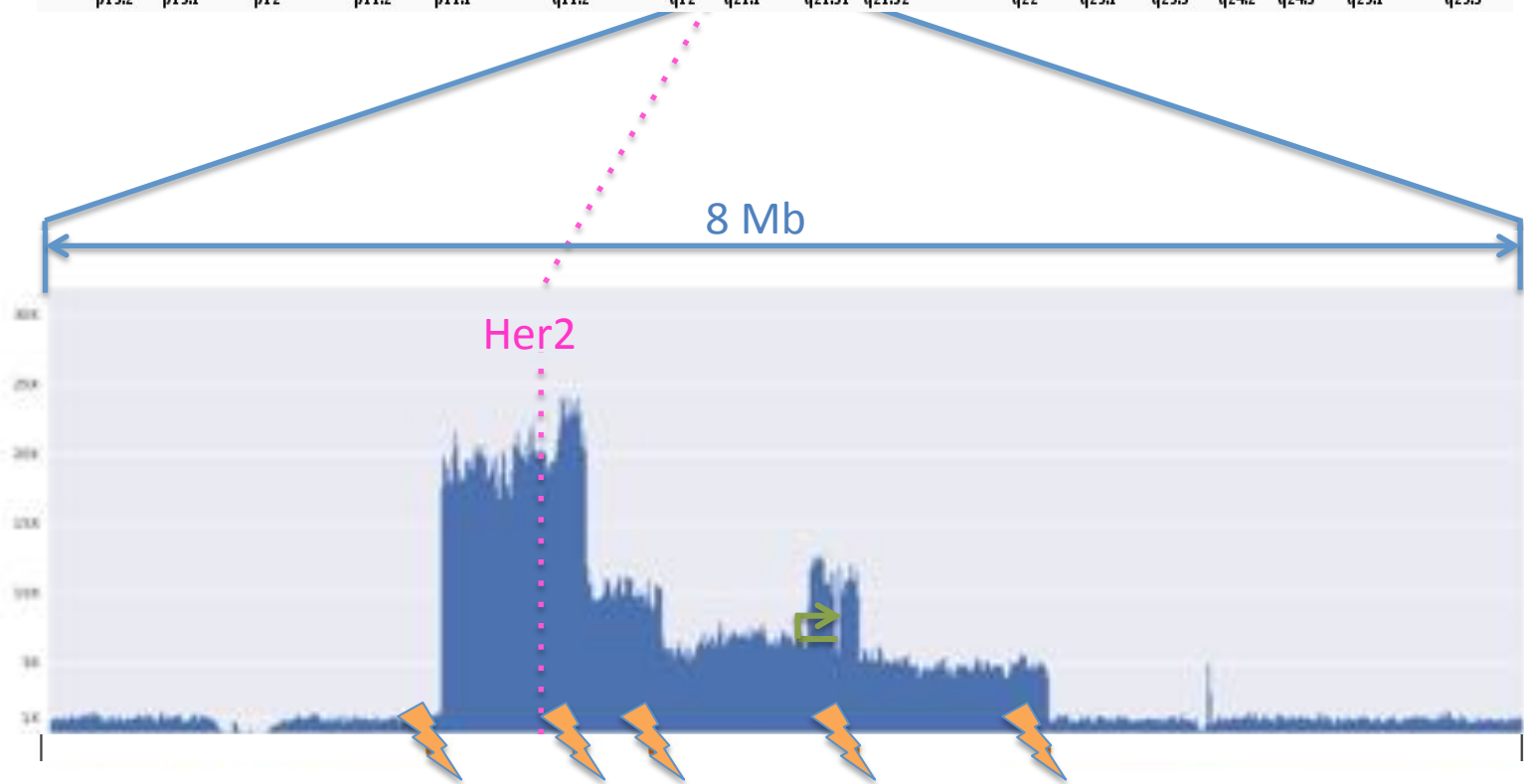


← Chr 17: 83 Mb →

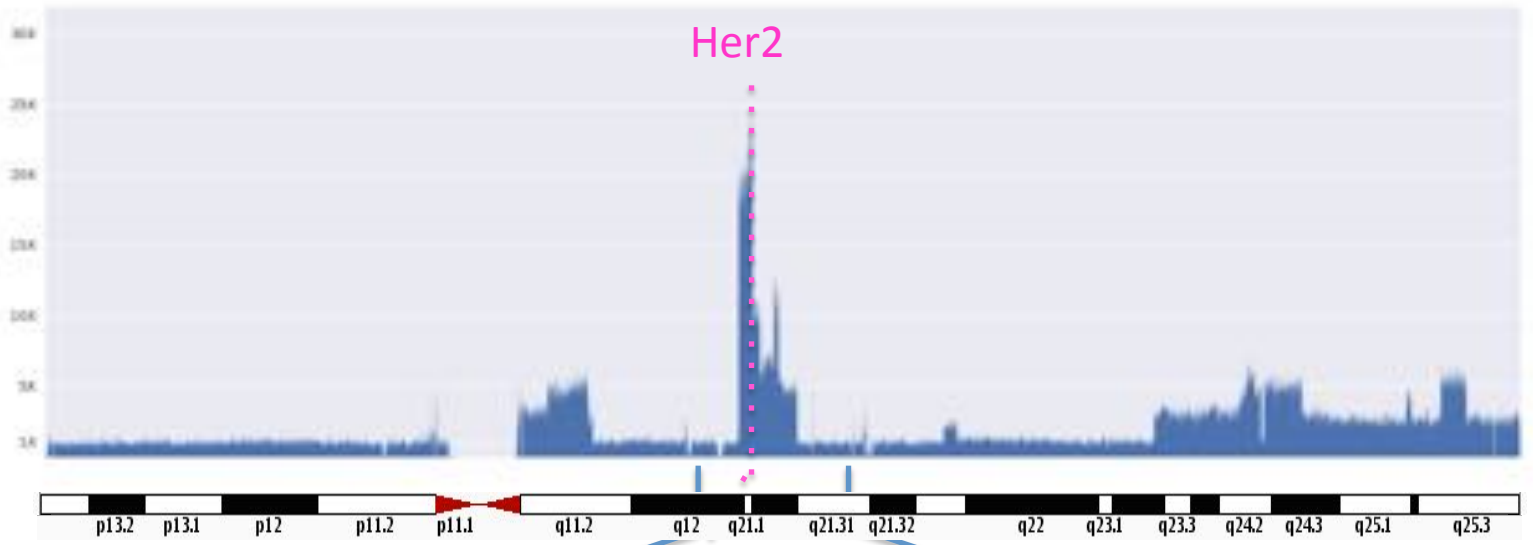
PacBio



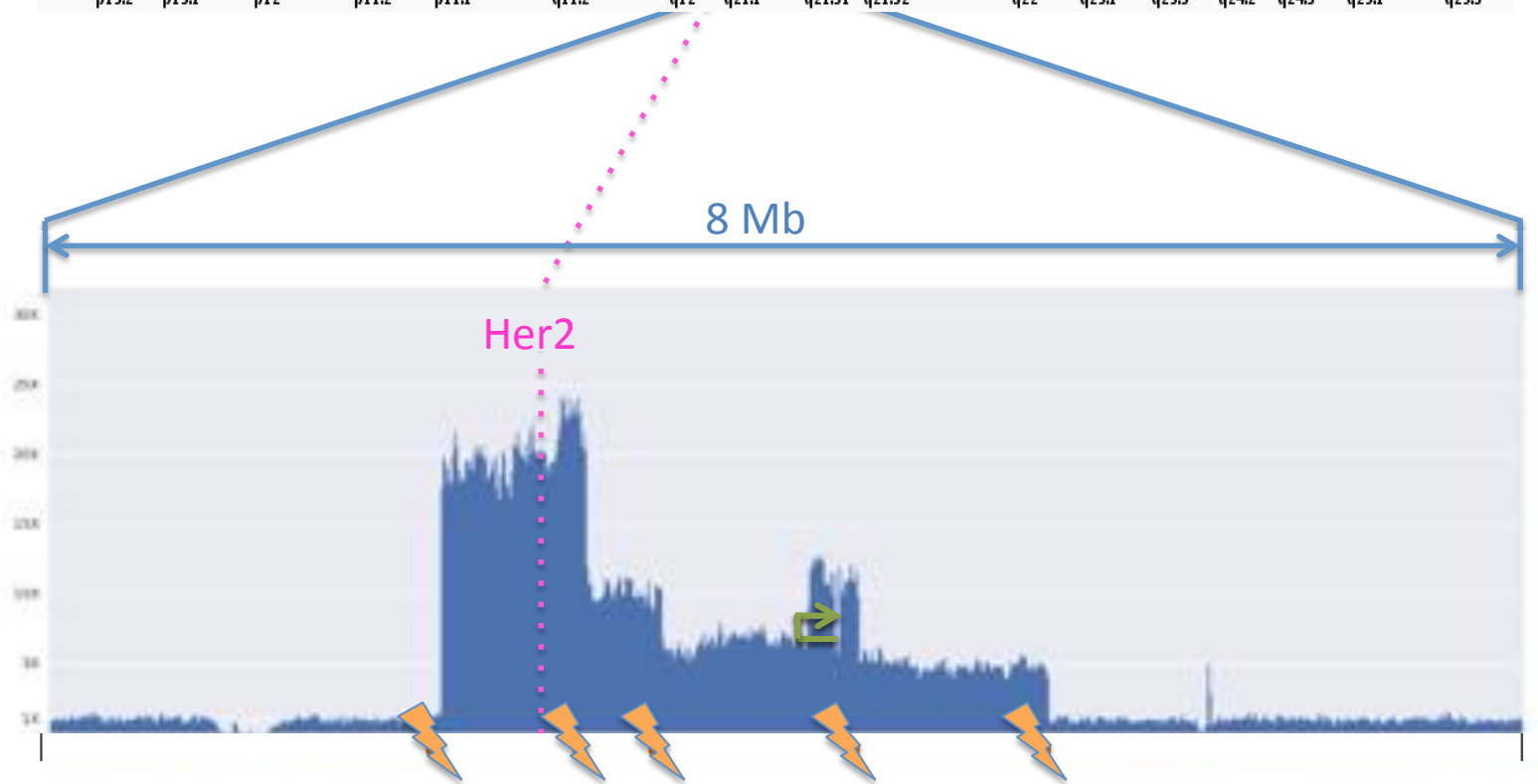
PacBio
chr17

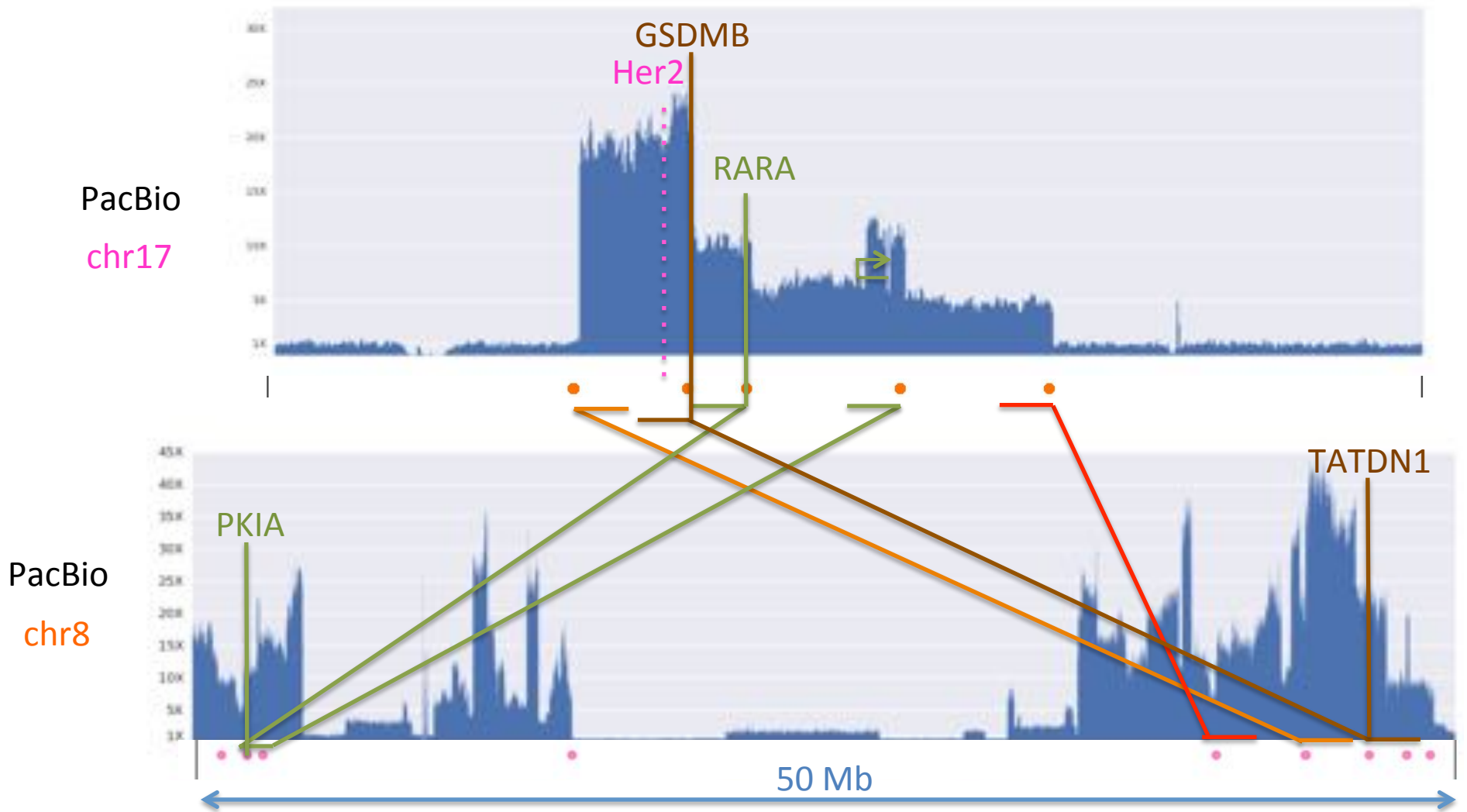


PacBio

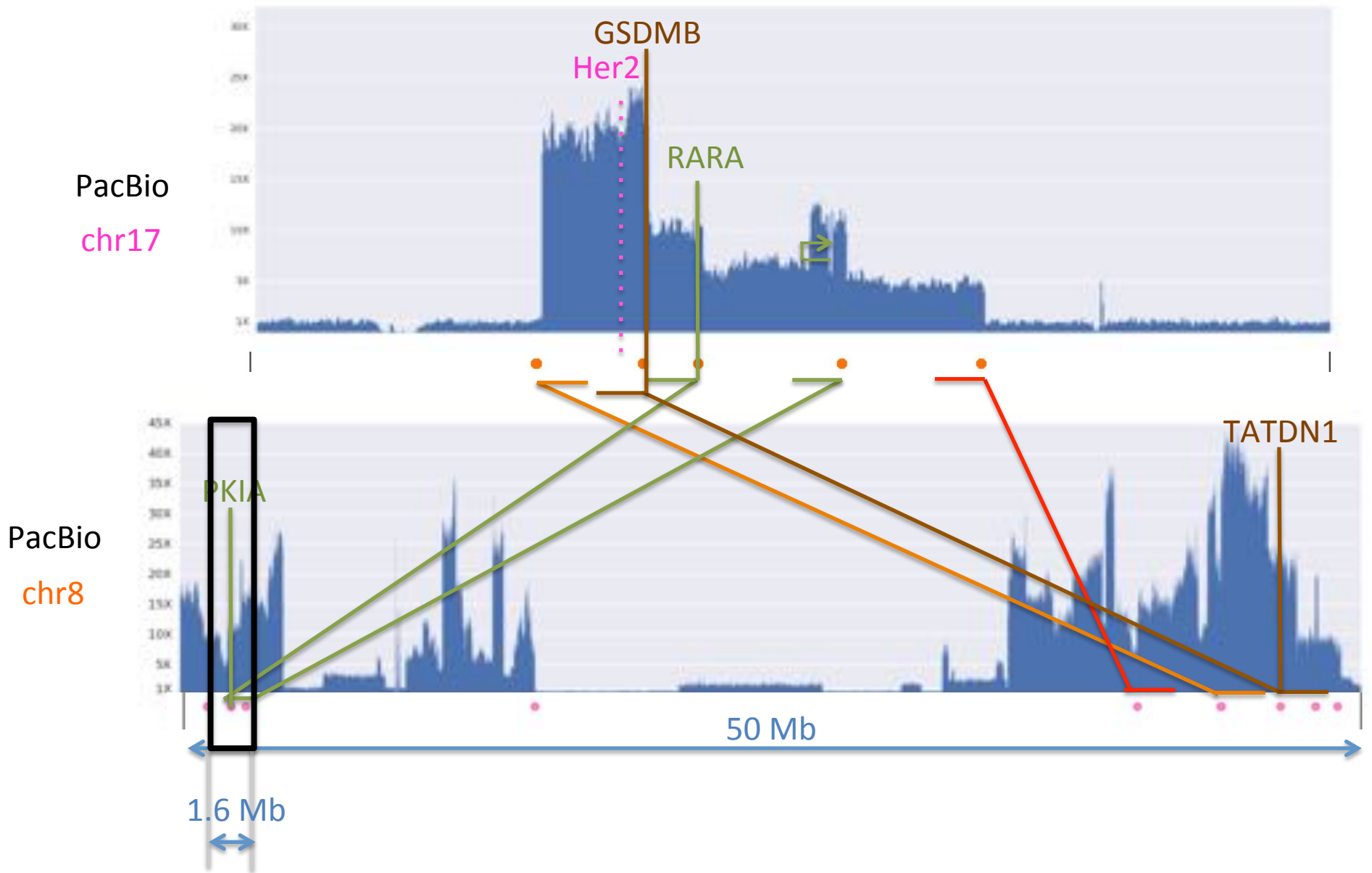


PacBio
chr17



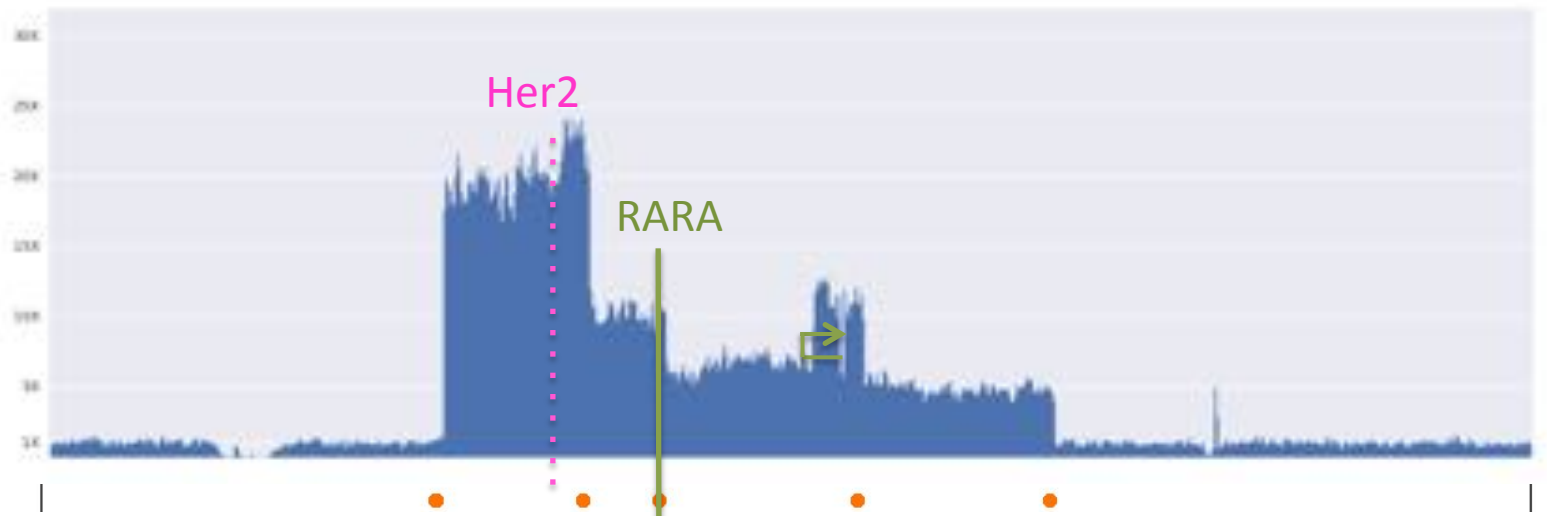


Confirmed both known gene fusions in this region

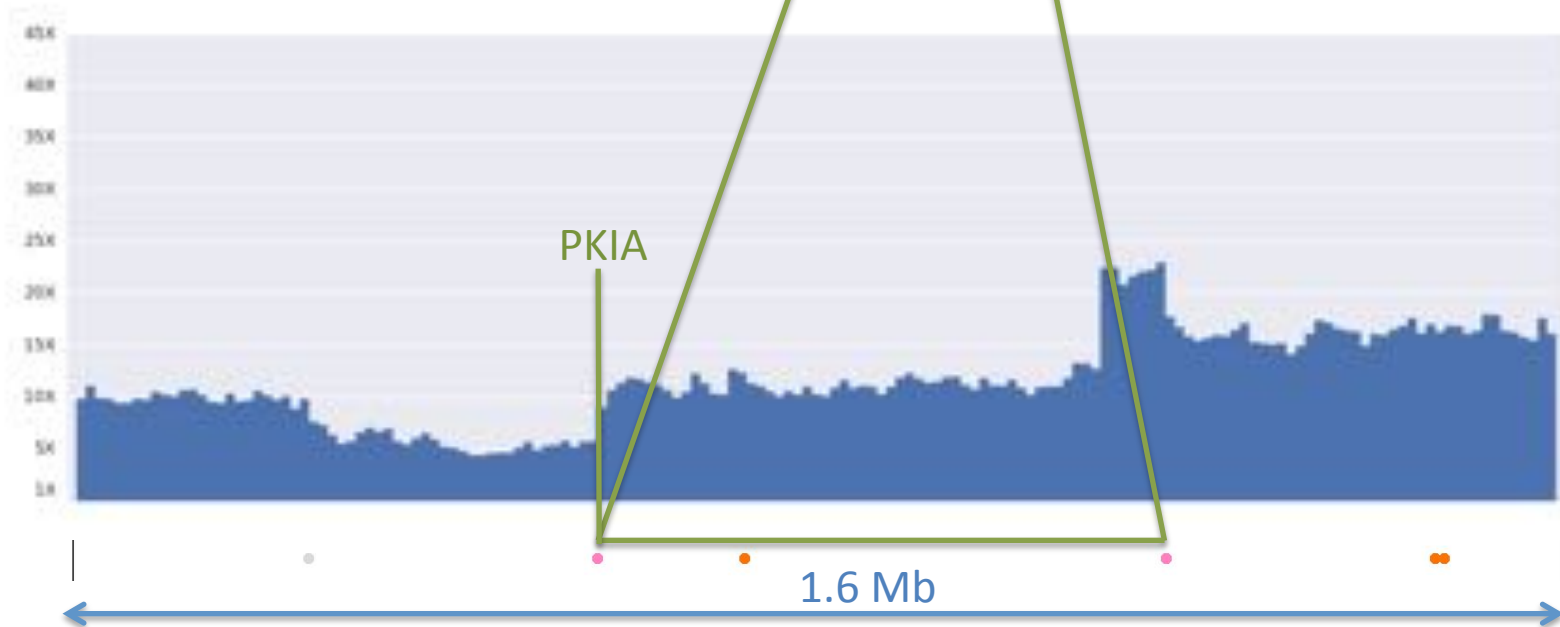


Confirmed both known gene fusions in this region

PacBio
chr17

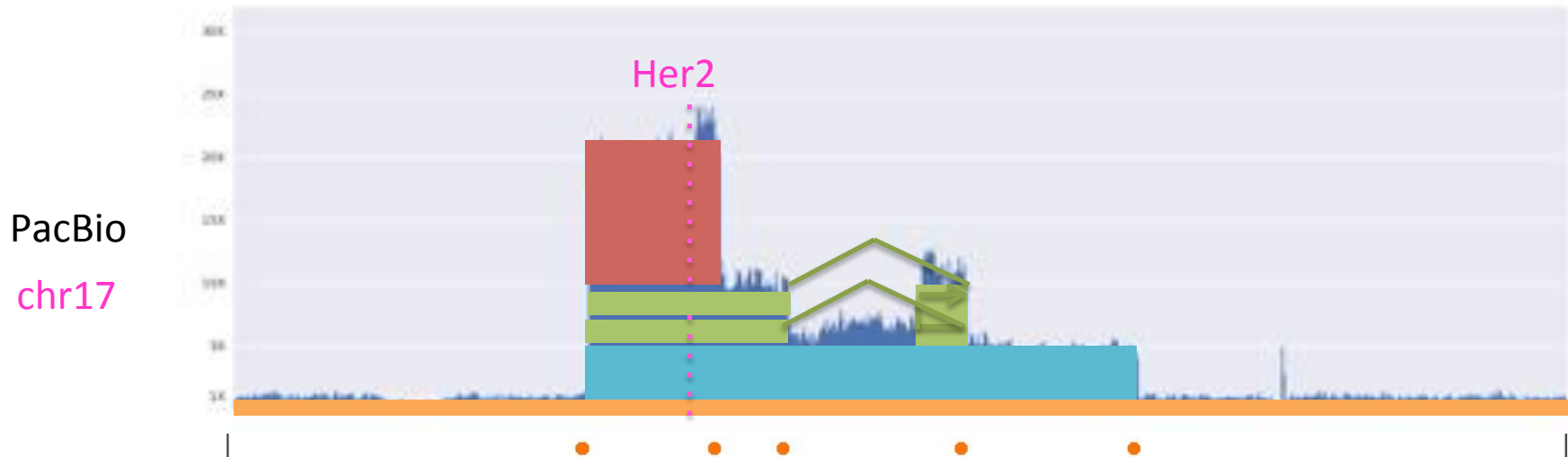


chr8



Joint coverage and breakpoint analysis to discover underlying events

Cancer lesion Reconstruction



By comparing the proportion of reads that are spanning or split at breakpoints we can begin to infer the history of the genetic lesions.

1. Healthy diploid genome
2. Original translocation into chromosome 8
3. Duplication, inversion, and inverted duplication within chromosome 8
4. Final duplication from within chromosome 8

Cancer lesion Reconstruction

Available *today* under the Toronto Agreement:

- Fastq & BAM files of aligned reads
- Interactive Coverage Analysis with BAM.IOBIO
- Whole genome assembly & alignment

Available soon

- Whole genome methylation analysis
- Full length cDNA transcriptome analysis
- Comparison to single cell analysis of >100 individual cells

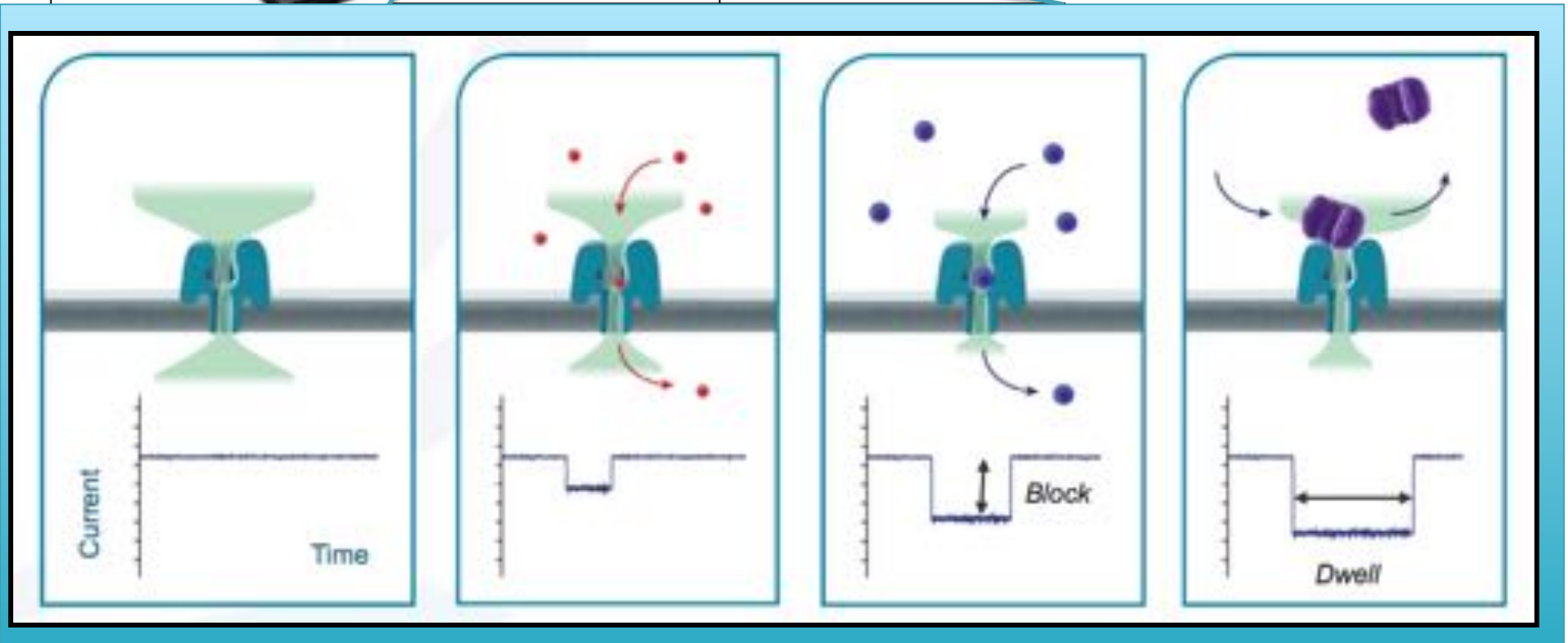
<http://schatzlab.cshl.edu/data/skbr3/>

4. Final duplication from within chromosome 8

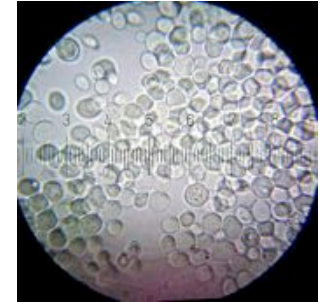
Oxford Nanopore MinION



- Thumb drive sized sequencer powered over USB
- Capacity for 512 reads at once
- Senses DNA by measuring changes to ion flow



Nanopore Readlengths



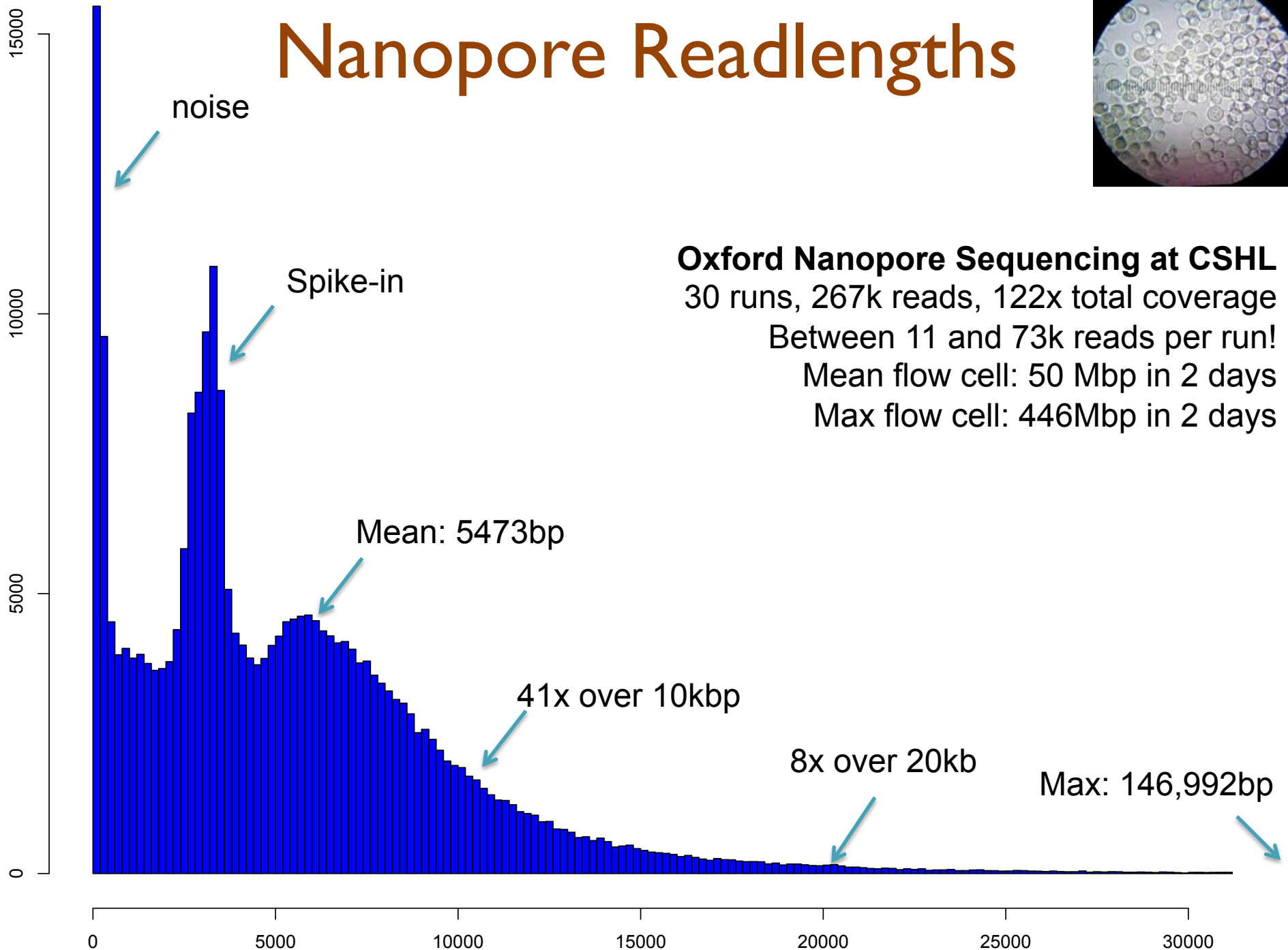
Oxford Nanopore Sequencing at CSHL

30 runs, 267k reads, 122x total coverage

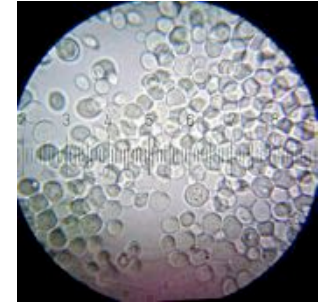
Between 11 and 73k reads per run!

Mean flow cell: 50 Mbp in 2 days

Max flow cell: 446Mbp in 2 days



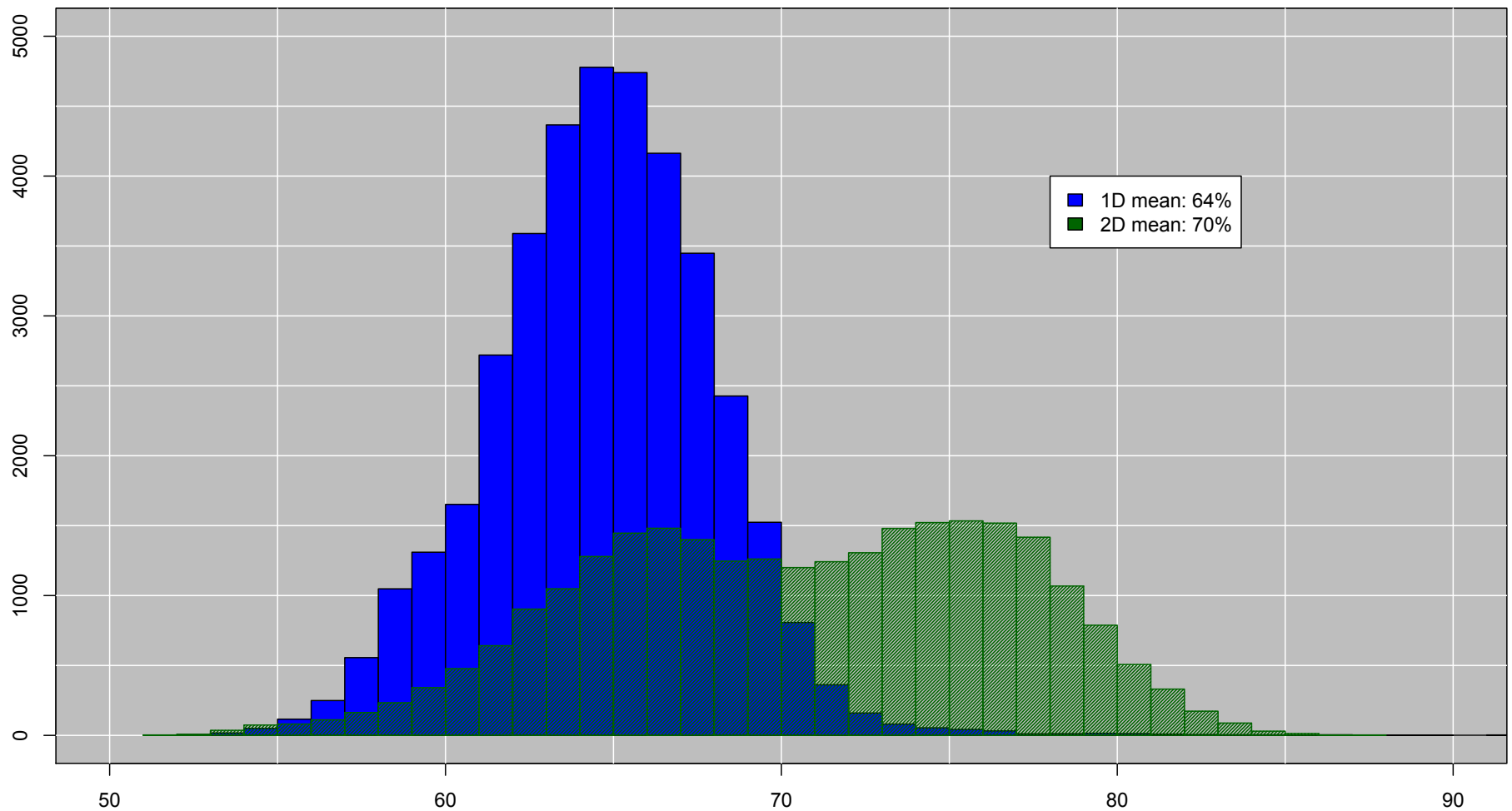
Nanopore Accuracy



Alignment Quality (BLASTN)

Of reads that align, average ~64% identity

“2D base-calling” improves to ~70% identity

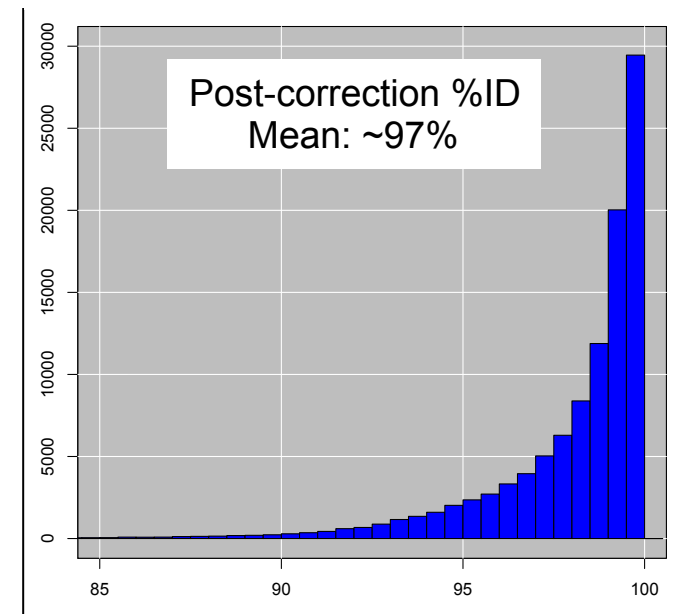
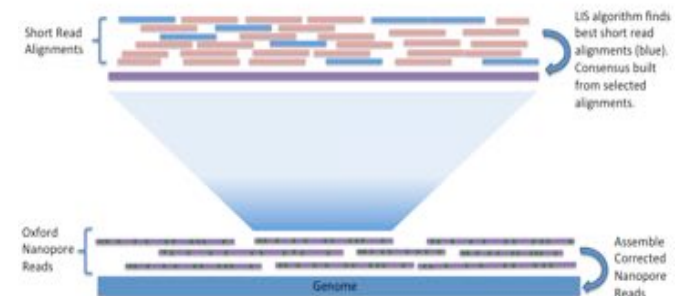


NanoCorr: Nanopore-Illumina Hybrid Error Correction



<https://github.com/jgurtowski/nanocorr>

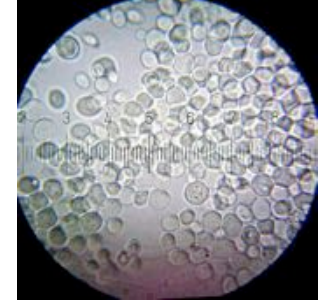
1. BLAST Miseq reads to all raw Oxford Nanopore reads
 - First pass scans to remove “contained” alignments
 - Second pass uses Dynamic Programming (LIS) to select set of high-identity alignments with minimal overlaps
2. Select non-repetitive alignments
 - State machine of most commonly observed base at each position in read
3. Compute consensus of each Oxford Nanopore read



Oxford Nanopore Sequencing and de novo Assembly of a Eukaryotic Genome

Goodwin, S, Gurtowski, J *et al.* (2015) bioRxiv doi: <http://dx.doi.org/10.1101/013490>

NanoCorr Yeast Assembly



S288C Reference sequence

- 12.1Mbp; 16 chromo + mitochondria; N50: 924kbp

bioRxiv
beta

THE PREPRINT SERVER FOR BIOLOGY

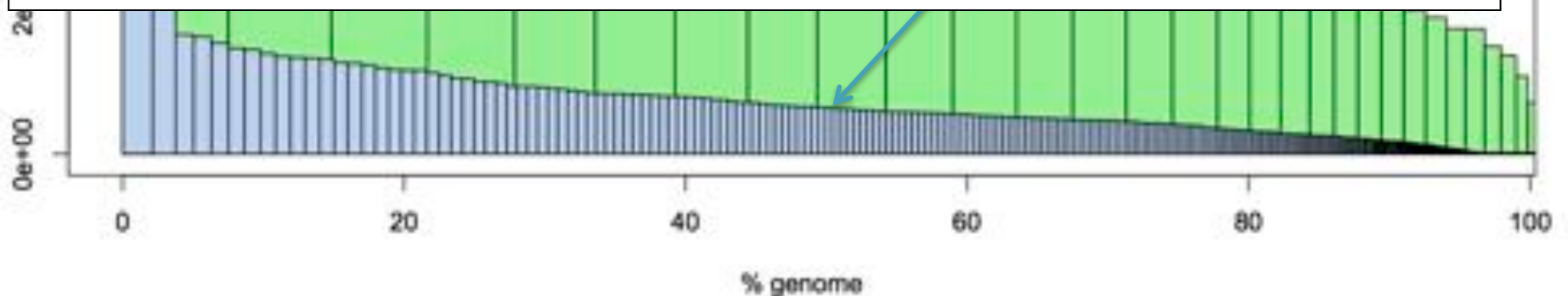
New Results

Oxford Nanopore Sequencing and de novo Assembly of a Eukaryotic Genome

Sara Goodwin , James Gurtowski , Scott Ethe-Sayers , Panchajanya Deshpande , Michael Schatz , W Richard McCombie

doi: <http://dx.doi.org/10.1101/013490>

contig length



Genomic Futures?

Zamin Iqbal and 5 others retweeted



GenomeWeb InSequence @InSequence · Oct 20

Oxford Nanopore shows off **PromethION** at **ASHG**. #ASHG14 #nanopore



Genomic Futures?



iGenomics: Mobile Sequence Analysis

Aspyn Palatnick, Elodie Ghedin, Michael Schatz

The worlds first genomics analysis app for iOS devices

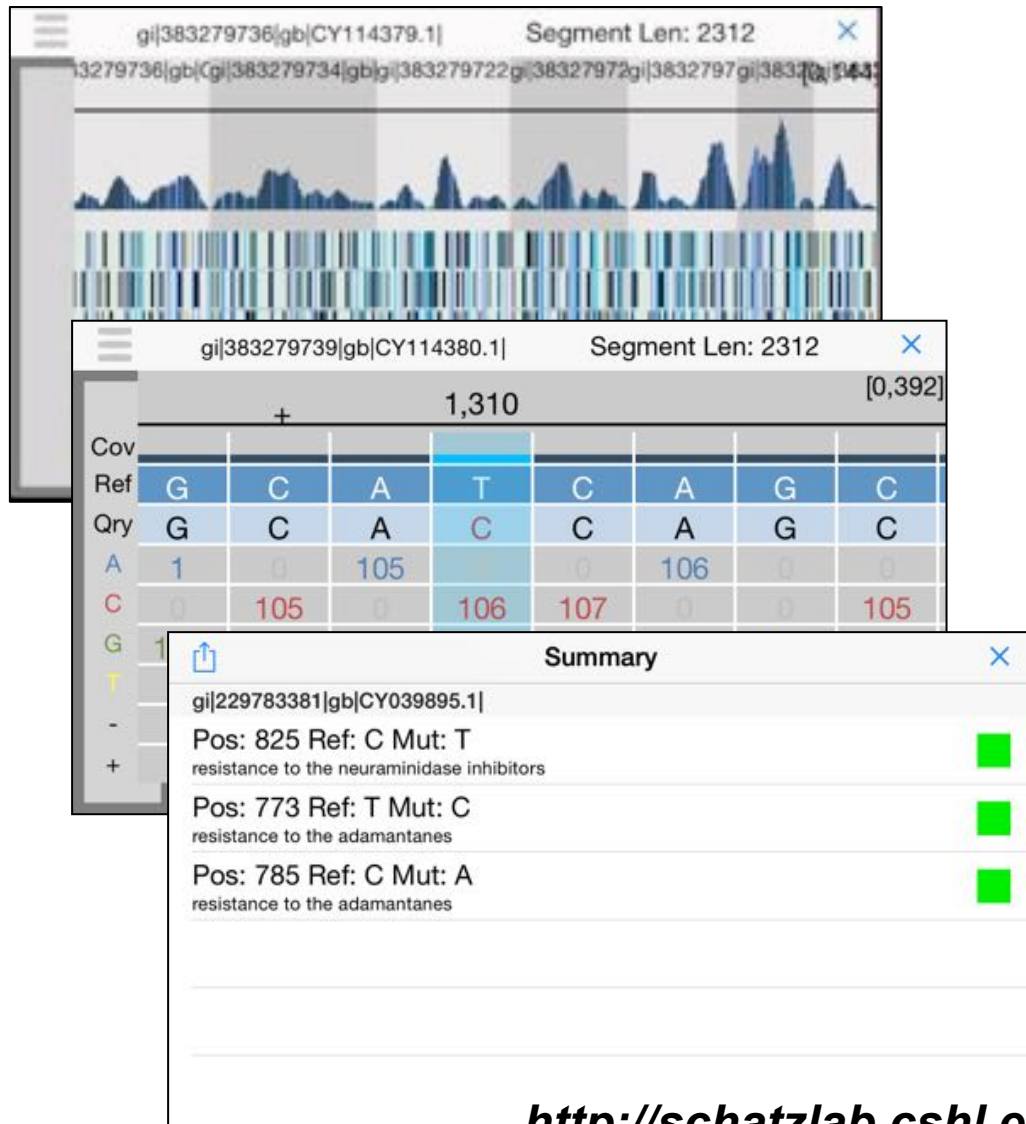
BWT + Dynamic Programming + UI

First application:

- Handheld diagnostics and therapeutic recommendations for influenza infections
- In the iOS AppStore now!

Future applications


- Pathogen detection
- Food safety
- Biomarkers
- etc..



<http://schatzlab.cshl.edu/iGenomics>

What should we expect from an assembly?

Summary & Recommendations



New Results

Error correction and assembly complexity of single molecule sequencing reads.

Hayan Lee , James Gurtowski , Shinjae Yoo , Shoshana Marcus , W. Richard McCombie , Michael Schatz
doi: <http://dx.doi.org/10.1101/006395>

> 5GB: Email mschatz@cshl.edu

The year 2015 will mark the return to reference quality genome sequence

technologies are quickly improving, exciting new scaffolding technologies

Acknowledgements

Schatz Lab

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Ramakrishnan
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Rachel Sherman
Greg Vulture
Alejandro Wences

CSHL

Hannon Lab
Gingeras Lab
Jackson Lab
Hicks Lab
Iossifov Lab
Levy Lab
Lippman Lab
Lyon Lab
Martienssen Lab
McCombie Lab
Tuveson Lab
Ware Lab
Wigler Lab

SBU

Skiena Lab
Patro Lab

Cornell

Susan McCouch
Lyza Maron
Mark Wright

OICR

John McPherson
Karen Ng
Timothy Beck
Yogi Sundaravadanam

NYU

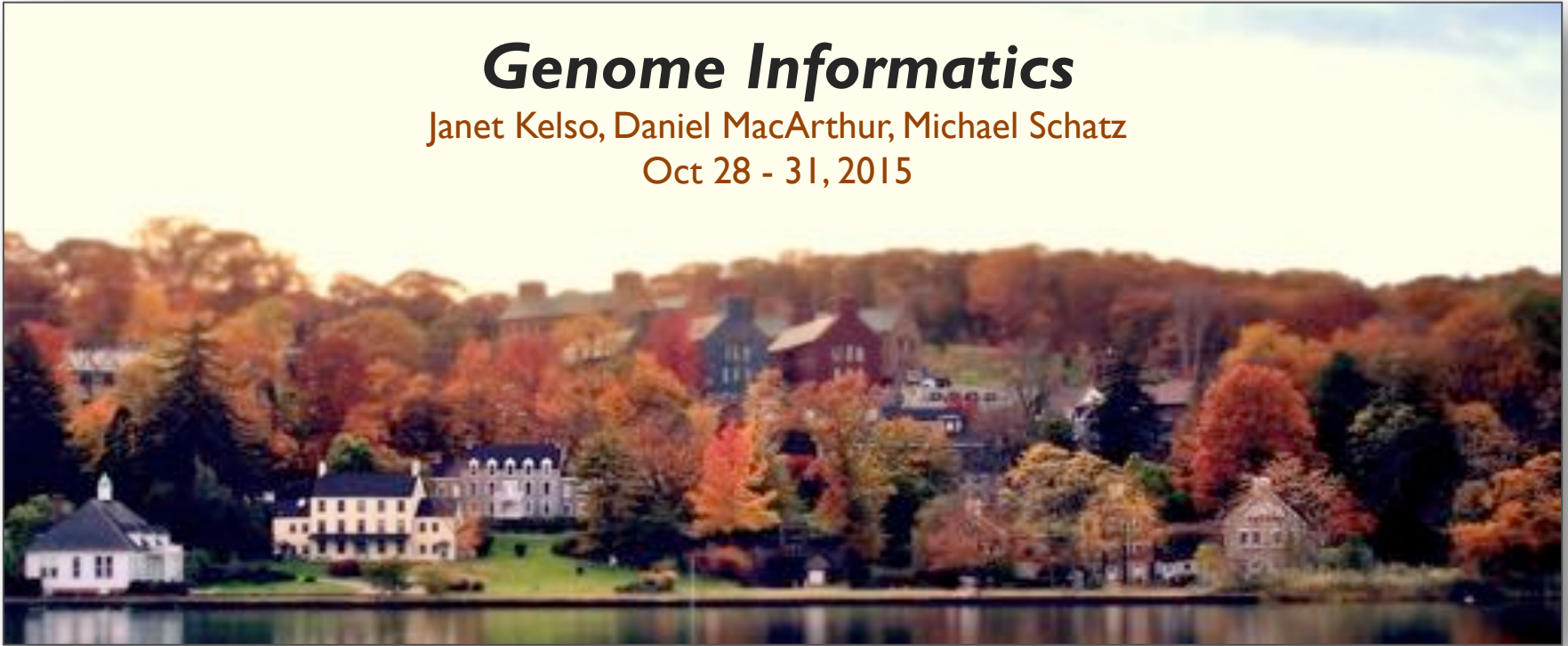
Jane Carlton
Elodie Ghedin



Genome Informatics

Janet Kelso, Daniel MacArthur, Michael Schatz

Oct 28 - 31, 2015



Thank you

<http://schatzlab.cshl.edu>

@mike_schatz